

Unconjugated Estriol EIA



AL-138-i

INTENDED USE

The Unconjugated Estriol (uE3) Enzyme Immunoassay (EIA) Kit provides materials for the quantitative measurement of unconjugated estriol in serum. It is intended for in vitro diagnostic use as an aid in the diagnosis and monitoring of various hormonal reproductive disorders. Down syndrome (Trisomy 21) screening in the 2nd trimester using combined biochemical and ultrasound markers may be assessed with appropriate risk calculation software.^{1,2} It is strongly recommended to use validated software (CE marked) designated specifically for evaluating the risk of Trisomy 21 using uE3 in combination with AFP, hCG, and Inhibin A measurements.

SUMMARY AND EXPLANATION

Estriol (1,3,5(10)-estratriene-3,16 α ,17 β -triol; E₃) is one of the 3 major naturally occurring estrogens, the others being estradiol and estrone. Estriol is produced almost exclusively during pregnancy, and is the major estrogen produced in the normal human fetus. During pregnancy, the production of estriol depends on an intact maternal-placental-fetal unit^{3,4}. Steroid precursors from the maternal circulation are taken up by the placenta and converted to progesterone. Progesterone is then converted to dehydroepiandrosterone sulfate (DHEA-S) in the fetal adrenal gland, which is then 16 α -hydroxylated in the fetal liver. 16 α -hydroxylase activity is present in only very low amounts in placenta and non-fetal tissues. In the placenta, 16 α -hydroxy-DHEA-S is then converted sequentially to 16 α -hydroxy-DHEA, 16 α -hydroxyandrost-4-ene-3,17-dione and, finally, estriol. Estriol may also originate from estrogen precursors, such as 16 α -hydroxyestrone. This pathway may account for the high levels of estrone sulfate found in breast cyst fluid.⁵

Fetal-placental production of estriol leads to a progressive rise in maternal circulating estriol levels, reaching a late-gestational peak which is ~2-3 orders of magnitude greater than non-pregnant levels^{3,4}. In the maternal circulation, estriol undergoes rapid conjugation in the liver followed by urinary excretion with a half-life of ~20 minutes³. Therefore, maternal estriol levels can provide a dynamic estimate of fetal production rates. In terms of estrogenic activity, estriol is much less potent than estradiol⁴. The physiologic role of estriol is not known.

Specific diagnostic and therapeutic uses for estriol measurements are not completely defined, although clinical utility during pregnancy has been investigated. Since normal estriol production depends on an intact maternal-placental-fetal circulation and functional fetal metabolism, maternal estriol levels have been used to monitor fetal status during pregnancy, particularly during the third trimester. Because estriol concentrations are subject to diurnal and episodic variation, it is common practice to refer serum measurements to a baseline, defined for the patient as either the average or the highest of her three most recent estriol results. However, studies in diabetic pregnancies suggest that low estriol levels have limited value in predicting fetal distress⁶. The AL-138 Unconjugated Estriol EIA uses a rabbit anti-estriol antibody preparation with low cross-reactivity to other natural estrogens and estrogen precursors.

PRINCIPLE OF THE TEST

The Unconjugated Estriol EIA Kit uses the competitive binding enzyme immunoassay format. In the assay, Calibrators, Controls and Unknowns containing unconjugated estriol are incubated with biotin-labeled estriol and rabbit anti-estriol antiserum in microtitration wells coated with goat anti-rabbit gamma globulin where the unlabeled and biotin-labeled antigens compete for a limited number of anti-estriol binding sites. After incubation and washing, the wells are incubated with streptavidin-horse radish peroxidase, which binds to the biotinylated estriol. The unbound streptavidin-HRP is washed, followed by incubation with the substrate tetramethylbenzidine (TMB). An acidic stopping solution is then added and the degree of enzymatic turnover of the substrate is determined by dual wavelength absorbance measurement at 450 and 620 nm.

MATERIALS SUPPLIED

CAL-138A - CAL-138F Estriol Calibrators

One vial each of 3.5 mL of Calibrator A and 0.5 ml of Calibrators B-F, containing concentrations of approximately 0 – 19.0 ng/mL Estriol in Estriol calibrator matrix. Refer to **calibration card** for exact concentrations. **Calibrators are shipped ambient. Store at - 20°C upon receipt until the expiration date.** Aliquot and freeze for multiple use. Avoid repeated freeze thaws.

Note: Estriol concentration in the Unconjugated Estriol EIA is traceable to the manufacturer's working calibrators.

CYB-138-I and CYB-138-II Estriol Controls

Two vials, 0.5 mL each, labeled Levels I and II containing low and high Estriol in Estriol calibrator matrix. Refer to **calibration card** for exact concentrations. **Controls are shipped ambient. Store at - 20°C upon receipt until the expiration date.** Aliquot and freeze for multiple use. Avoid repeated freeze thaws.

AG002 Goat Anti-Rabbit IgG (GARG) Microtitration Strips

One stripholder containing 96 polystyrene microtitration wells with Goat anti-rabbit IgG antibody immobilized to the inside wall of each well. Store at 2 to 8°C until expiration date in the resealable pouch with a desiccant to protect from moisture.

BCC-138 Estriol Biotin Conjugate Concentrate (50X)

One bottle, 0.4 mL, containing a protein-based buffer with a non-mercury preservative. Store at 2 to 8°C until expiration date.

CND-138 Estriol Biotin Conjugate Diluent

One bottle, 12 mL, containing a protein-based buffer with a non-mercury preservative. Store at 2 to 8°C until expiration date.

ABS-138 Anti-Estriol Antibody Solution

One bottle, 12 mL, containing anti-Estriol rabbit polyclonal antibody in a protein-based buffer with a non-mercury preservative. Store at 2 to 8°C until expiration date.

SAR-138 Estriol Streptavidin Conjugate Ready-to-Use

One bottle, 12 mL, containing streptavidin-HRP (horseradish peroxidase) in a protein-based buffer and a non-mercury preservative. Store undiluted at 2-8°C until expiration date.

TMB-100 TMB Chromogen Solution

One bottle, 12 mL, containing a solution of tetramethylbenzidine (TMB) in buffer with hydrogen peroxide. Store at 2 to 8°C until expiration date.

WSH-100 Wash Concentrate A

One bottle, 60 mL, containing buffered saline with a nonionic detergent. Store at 2 to 30°C until expiration date. Dilute 25-fold with deionized water prior to use.

STP-100 Stopping Solution

One bottle, 12 mL, containing 0.2 M sulfuric acid. Store at 2 to 30°C until expiration date.

MATERIALS REQUIRED BUT NOT PROVIDED

1. Microtitration plate reader capable of absorbance measurement at 450 nm, 405nm and 630 nm.
2. Microtitration orbital plate shaker
3. Microtitration plate washer.
4. Semi-automated/manual precision pipette to deliver 10-250 μ L.
5. Repeater pipette
6. Repeater plus for repeated dispensing
7. Vortex mixer.
8. Deionized water.

WARNINGS AND PRECAUTIONS**For *in vitro* diagnostic use.**

The following precautions should be observed:

- a) Follow good laboratory practice.
- b) Use personal protective equipment. Wear lab coats and disposable gloves when handling immunoassay materials.
- c) Handle and dispose of all reagents and material in compliance with applicable regulations.

WARNING: Potential Biohazardous Material

This reagent may contain some bovine and rabbit antiserum, or materials used in conjunction with animal source materials. Handle all reagents and patient samples at a Biosafety Level 2, as recommended for any potentially infectious human material in the Centers for Disease Control/National Institutes of Health manual "Biosafety in Microbiological and Biomedical Laboratories," 6th Edition, 2020⁷.

WARNING: Potential Chemical Hazard

Some reagents in this kit contain ProClin™ 300 as a preservative. ProClin™ 300 in concentrated amounts is an irritant to skin and mucous membranes. For further information regarding hazardous substances in the kit, please refer to the SDS, either at AnshLabs.com or by request.

SAMPLE COLLECTION AND PREPARATION

- Serum is the recommended sample type.
- Sample handling, processing, and storage requirements depend on the brand of blood collection tube⁸ that you use. Please reference the manufacturer's instructions for guidance. Each laboratory should

determine the acceptability of its own blood collection tubes and serum separation products.

- Avoid assaying lipemic, hemolyzed or icteric samples.
- Avoid repeated freezing and thawing of samples.
- For shipping, place specimens in leak proof containers in biohazard specimen bags with appropriate specimen identification and test requisition information in the outside pocket of the biohazard specimen bag. Follow DOT and IATA requirements when shipping specimens.

PROCEDURAL NOTES

1. A thorough understanding of this package insert is necessary for successful use of the Estriol EIA assay. It is the responsibility of the customer to validate the assay for their use. Accurate results will only be obtained by using precise laboratory techniques and following the package insert.
2. A calibration curve must be included with each assay.
3. Bring all kit reagents to room temperature before use. Thoroughly mix the reagents before use by gentle inversion. Do not mix different lots of any kit component and do not use any component beyond the expiration date.
4. Use a clean disposable pipette tip for each reagent, calibrator, control, or sample. Avoid microbial contamination of reagents, contamination of the substrate solutions with the SHRP conjugate. The enzyme used as the label is inactivated by oxygen, and is highly sensitive to microbial contamination, sodium azide, hypochlorous acid and aromatic chlorohydrocarbons often found in laboratory water supplies. Use deionized water.
5. Incomplete washing will adversely affect the outcome and assay precision. Care should be taken to add TMB into the wells accurately and efficiently to minimize potential assay drift due to variation in the TMB incubation time. Avoid exposure of the reagents to excessive heat or direct sunlight.

PREPARATION OF REAGENTS

1. **Wash Solution:** Dilute wash concentrate 25-fold with deionized water. The wash solution is stable for one month at room temperature when stored in a tightly sealed bottle.
2. **Microtitration Wells:** Select the number of coated wells required for the assay. The remaining unused wells should be placed in the resealable pouch with a desiccant. The pouch must be resealed to protect from moisture.
3. **Estriol Biotin Conjugate Solution:** The Estriol Biotin Conjugate Concentrate should be diluted at a ratio of 1 part conjugate to 50 parts of Estriol Biotin Conjugate Diluent according to the number of wells used. If an entire plate is to be used pipet exactly 220 μ L of the Concentrate into 11 mL of the diluent.
Note: Estriol biotin conjugate concentrate should be freshly diluted 10 minutes prior to use.

ASSAY PROCEDURE

Allow all specimens and reagents to reach room temperature ($23 \pm 2^\circ\text{C}$) and mix thoroughly by gentle inversion before use. Calibrators, controls, and unknowns should be assayed in duplicate.

NOTE: All serum samples reading higher than the highest calibrator should be mixed and diluted in the 0 ng/mL Calibrator A prior to assay.

1. Prepare the **Estriol-Biotin Conjugate Solution** by diluting the Estriol Biotin Conjugate Concentrate in Estriol Conjugate Diluent as described under the Preparation of the Reagents section
2. Label the microtitration strips to be used.
3. Pipette 50 μ L of the **Calibrators** and **Controls** to the appropriate wells.

4. Pipette **25 µL** of the **Calibrator A** to the remaining wells which will be used. Pipette **25 µL** of **Unknowns** in the wells containing Calibrator A.
5. Add **100 µL** of **Estriol-Biotin Conjugate Solution** to each well using a repeater pipette.
6. Add **100 µL** of **Estriol Antibody Solution** to each well using a repeater pipette.
7. Incubate the plate, shaking at a fast speed (**600-800 rpm**) on an orbital microplate shaker, for **30 minutes** at room temperature ($23 \pm 2^\circ\text{C}$).
8. Aspirate and wash each strip 5 times with Washing Solution (350 µL/per well) using an automatic microplate washer.
9. Add **100 µL** of the **Streptavidin-Enzyme Conjugate-RTU** to each well using a repeater pipette.
10. Incubate the plate, shaking at a fast speed (**600-800 rpm**) on an orbital microplate shaker, for **30 minutes** at room temperature ($23 \pm 2^\circ\text{C}$).
11. Aspirate and wash each strip 5 times with the Wash Solution (350 µL/per well) using an automatic microplate washer.
12. Add **100 µL** of the **TMB chromogen solution** to each well using a repeater pipette. Avoid exposure to direct sunlight.
13. Incubate the wells, shaking at **600-800 rpm** on an orbital microplate shaker, for **10-12 min** at room temperature ($23 \pm 2^\circ\text{C}$).
NOTE: Visually monitor the color development to optimize the incubation time.
14. Add **100 µL** of the **Stopping solution** to each well using a repeater pipette. Read the absorbance of the solution in the wells within **20 minutes**, using a microplate reader set to **450 nm**.
NOTE: If instrument has a wavelength correction, set the instrument to dual wavelength measurement at **450 nm** with background wavelength correction at **630 nm**.

RESULTS

1. Optimum results can be obtained at incubation temperature of $23 \pm 2^\circ\text{C}$.
2. Calculate the mean OD for each calibrator, control, and sample.
3. Plot the mean OD readings for each of the Calibrators along the y-axis versus log of Estriol calibrator concentrations in ng/mL on the x-axis, using a 4PL or sigmoid curve fit.
4. Determine the Estriol concentrations of the controls and samples from the calibration curve by matching their mean OD readings with the corresponding Estriol concentrations.
5. Any sample reading higher than the highest calibrator should be appropriately diluted into a low reading serum sample and re-assayed.
6. Any sample reading lower than the analytical sensitivity should be reported as such.

LIMITATIONS

The reagents supplied in this kit are optimized to measure Estriol levels in Estriol calibrator matrix. If there is evidence of microbial contamination or excessive turbidity in a reagent, discard the vial. For assays employing antibodies, the possibility exists for interference by heterophile antibodies in the samples⁹.

As with all diagnostic tests, a definite clinical diagnosis **should NOT** be based on the results of a **single test**. The diagnosis should only be made by the physician after reviewing all clinical and laboratory findings.

QUALITY CONTROL

- Each laboratory should establish mean values and acceptable ranges to ensure proper performance.
- Estriol EIA controls or other commercial controls should fall within established confidence limits.

- The confidence limits for Estriol EIA controls are printed on the calibration card.
- A full calibration curve, low- and high-level controls, should be included in each assay.
- The TMB chromogen solution should be colorless. Development of a blue color may indicate reagent contamination or instability.

REPRESENTATIVE CALIBRATION CURVE DATA

Well Number	Well Contents Calibrators	Mean Absorbance	Conc (ng/mL)
A1, A2	A	3.394	0
B1, B2	B	2.820	0.054
C1, C2	C	1.953	0.23
D1, D2	D	1.066	0.84
E1, E2	E	0.629	2.3
F1, F2	F	0.306	11.75

CAUTION: The above data must not be employed in lieu of data obtained by the user in the laboratory.

ANALYTICAL CHARACTERISTICS

All analytical characteristics are stated ng/mL.

Conversion Factor: ng/mL x 3.5 = nmol/L

Analytical Sensitivity:

The analytical sensitivity in the assay as calculated by the interpolation of mean plus two standard deviation of 16 replicates of calibrator A (0 ng/mL) and calibrator B (0.053 ng/mL) is 0.04 ng/mL.

Limit of Blank (LOB):

The Limit of Blank is 0.006 ng/mL, calculated from a minimum of n=16 measurements of analyte free sample.

Limit of Detection (LOD):

The lowest amount of Estriol in a sample that can be detected with a 95% probability (n=12) is 0.05 ng/mL. The value was determined by processing five serum samples in the range of 0.05 ng/mL to 0.8 ng/mL.

Limit of Quantitation (LOQ):

The estimated minimum dose achieved at 20% total imprecision is 0.03 ng/mL. The value was determined by processing five samples in the range of 0.05 ng/mL to 0.8 ng/mL over four runs and two days in triplicates (n=12).

Imprecision:

Reproducibility of the Estriol EIA assay was determined in a study using three serum pools. The study included a total of 5 assays, three replicates of each per assay (n=15). Representative data were calculated and are presented in the following table.

Sample	Mean Conc.	Within Run		Between Run		Total	
		SD	%CV	SD	%CV	SD	%CV
	(ng/mL)						
Pool-1	0.75	0.04	5.80%	0.03	4.16%	0.05	7.13%
Pool-2	2.59	0.16	6.31%	0.15	5.68%	0.22	8.49%
Pool-3	7.22	0.77	10.65%	0.53	7.30%	0.93	12.91%

Recovery:

Known amounts of Estriol were added to five serum samples containing different levels of endogenous Estriol. The concentration of Estriol was

determined before and after the addition of exogenous Estradiol and the percent recovery was calculated.

Sample ID	Endogenous Value in ng/mL	Expected in ng/mL	Observed in ng/mL	%Recovery
S1	1.52	2.32	2.56	110%
		3.13	3.39	108%
		3.93	4.24	108%
S2	1.35	2.16	2.32	107%
		2.97	3.01	101%
S3	1.18	2.01	2.24	111%
		2.83	3.08	109%
		3.65	3.87	106%
S4	0.90	1.73	1.74	101%
		2.57	2.33	91%
S5	1.65	3.40	3.03	89%
		4.44	4.63	107%
		4.00	4.40	109%

Linearity:

Two serum samples containing various Estradiol levels and Calibrator F were diluted with calibrator A. The % recovery on individual samples is represented in the following table.

Sample ID	Dilution factor (1 in X)	Expected Value in ng/mL	Observed Value in ng/mL	Average % Recovery
Calibrator F	Neat Value	11.2	11.2	99%
	2	5.6	5.58	
	4	2.8	2.72	
	8	1.4	1.48	
	16	0.7	0.73	
	32	0.35	0.35	
S1	Neat Value	6.14	6.14	110%
	2	3.07	3.05	
	4	1.53	1.64	
	8	0.77	0.89	
	16	0.38	0.43	
	32	0.19	0.22	
S2	Neat Value	4.36	4.36	102%
	2	2.18	2.19	
	4	1.09	1.22	
	8	0.55	0.59	
	16	0.27	0.27	
	32	0.14	0.12	

Analytical Specificity:

The monoclonal antibody pair used in the assay detects Estradiol. Other related molecules at 100 ng/mL and their % cross-reactivity are represented in the following table.

Cross-Reactant	Concentration (ng/mL)	% Cross-Reactivity
Estrone	100	ND
Estradiol	100	0.822%
Estrone sulphate	100	ND
17- α -ethynyl estradiol	100	ND
β -Estradiol 17-(β -D-glucuronide) Sodium salt	100	0.045%
Aldosterone	100	ND
Androsterone	100	ND
Androstenediol	100	ND
DHEA	100	ND

Progesterone	100	ND
Testosterone	100	ND

ND= Non-Detectable (Less than LOD)

The monoclonal antibody pair used in the assay detects Goat, Rabbit, Bovine, Equine, Ovine, Mouse and Bovine and Equine follicular fluid and does not detect Porcine and Canine samples.

Interference:

When hemoglobin and triglycerides were added at a greater than two folds of their physiological concentration to control sample, Estradiol concentrations were within $\pm 20\%$ of the control as represented in the following table.

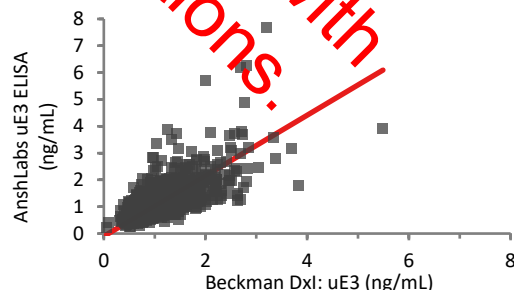
Interferent	Interferent Dose	Unspiked Sample (ng/mL)	Spiked Sample (ng/mL)	% Difference
Hemoglobin	1 mg/mL	6.01	5.42	-9.82
	0.5 mg/mL	6.35	6.53	2.83
	0.1 mg/mL	6.75	6.65	-1.39
Hemoglobin	1 mg/mL	2.87	3.19	11.01
	0.5 mg/mL	2.63	2.60	-0.93
	0.1 mg/mL	2.84	3.01	5.96
Biotin	1200 ng/mL	5.96	5.79	-2.84
	200 ng/mL	5.74	5.96	3.87
Biotin	1200 ng/mL	2.47	2.67	8.39
	200 ng/mL	3.21	3.29	2.56
Intralipids	20 mg/mL	5.24	4.97	-5.11
	10 mg/mL	5.58	5.81	4.16
	5 mg/mL	5.14	5.25	2.13
Intralipids	20 mg/mL	2.15	2.19	1.84
	10 mg/mL	2.32	2.25	-3.31
	5 mg/mL	2.62	2.46	-6.35
Bilirubin	0.66 mg/mL	3.13	3.31	5.54
	0.2 mg/mL	5.16	5.95	15.38
	0.1 mg/mL	5.86	5.53	-5.76
Bilirubin	0.66 mg/mL	1.76	1.95	11.16
	0.2 mg/mL	2.08	2.24	7.95
	0.1 mg/mL	2.38	2.38	0.19

Method Comparison:

The Estradiol EIA has been compared to a commercially available Estradiol EIA kit (Method A) using 830 second trimester pregnancy serum samples. Passing Bablok analysis of the results yielded the following regression:

$$\text{Estradiol EIA (AL-28)} = -0.1024 + 1.129 (\text{Beckman Dxl})$$

$$(r=0.7; P<0.0001)$$



Expected Value:

Serum samples were analyzed using Ansh Estradiol EIA. The expected ranges for Estradiol were calculated using 95% non-parametric estimation using Analyse-It® for Microsoft Excel.

Gestation Age	No. of Specimens (n)	Median Conc. (ng/mL)	Estradiol (ng/mL) 95% CI
15 weeks	229	0.77	0.36 – 1.54

16 weeks	259	0.94	0.43 – 2.42
17 weeks	158	1.16	0.56 – 2.80
18 weeks	108	1.4	0.65 – 3.18
19 weeks	48	1.91	0.68 – 4.85
20 weeks	26	1.98	1.12 – 3.78
21 weeks	12	2.1	1.37 – 4.89
22 weeks	9	2.79	1.25 – 7.67

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Les consommables pour Ansh Labs sont livrés avec des instructions d'utilisation en anglais. N'hésitez pas à contacter votre société d'assistance technique ou votre représentant Ansh Labs local pour obtenir des instructions traduites.

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FOR EXPORT ONLY

This assay is intended for *In Vitro Diagnostic Use*. **Not for sale in U.S.A**

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Symbols used with Ansh Labs Assays

Symbol	English	Deutsch	Français	Español	Italiano
	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instructions d'utilisation	Consulte las instrucciones de uso	Consultare le istruzioni per l'uso
	European Conformity	CE-Konformitätskennzeichnung	Conformité aux normes européennes	Conformidad europea	Conformità europea
	In vitro diagnostic device	In-vitro-Diagnostikum	Usage Diagnostic in vitro	Para uso Diagnóstico in vitro	Per uso Diagnostica in vitro
	Catalogue number	Katalog-Nr.	Numéro de catalogue	Número de catálogo	Numero di Catalogo
	Lot. No. / Batch code	Chargen-Nr.	Numéro de lot	Número de lote	Numero di lotto
	Storage Temperature	Lagerungstemperatur	Température de conservation	Temperatura de conservación	Temperatura di conservazione
	Expiration Date	Mindesthaltbarkeits-datum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
	Legal Manufacture	Hersteller	Fabricant	Fabricante	Fabbricante
Content	Content	Inhalt	Conditionnement	Contenido	Contenuto
Volume/No.	Volume / No.	Volumen / Anzahl	Volume/Quantité	Volumen/Número	Volume/Quantità

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